

Design of β -Hairpin Peptidomimetics That Inhibit Binding of α -Helical HIV-1 Rev Protein to the Rev Response Element RNA**

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The virally encoded Rev protein of human immunodeficiency virus type-1 (HIV-1) plays a critical role in viral replication by regulating the transport of unspliced and partially spliced viral RNA from the nucleus to the cytoplasm of infected cells.^[1,2] Rev binds first to a specific region of the HIV-1 mRNA, called the Rev response element (RRE) of stem loop IIb (Figure 1); subsequent to this initial binding event, approximately 10 additional Rev molecules oligomerize

through protein–protein and protein–RNA interactions and coat the entire RRE.^[3] Because of its essential role in viral replication, the interaction between Rev protein and the high-affinity RRE binding site represents an attractive yet unexploited target for antiviral therapy.

Rev recognition of the high-affinity RRE binding site is mediated by a small N-terminal 17-residue Arg-rich RNA-binding motif (Figure 1, bottom).^[4] NMR spectroscopic and biochemical studies have shown that the distorted A-form RRE RNA defines a deep binding pocket that is recognized by the peptide.^[5–9] This protein segment is unfolded in isolation, but adopts a α -helical conformation upon binding the RRE.^[6,9] To date, the search for drugs that target RRE has largely focused on derivatives of known aminoglycoside antibiotics, diphenylfurans, as well as related compounds that bind to the DNA minor groove,^[10] such as proflavines.^[11,12] However, this approach has failed to yield inhibitors of sufficient potency and selectivity to warrant further development.

Encouraged by recent success in using a β -hairpin structure to mimic an α -helical epitope in p53 and inhibit the p53-HDM2 interaction,^[13] we explore herein a new approach to inhibitors of the Rev–RRE interaction based on conformationally constrained β -hairpin peptidomimetics. We reasoned that a β -hairpin might provide a robust scaffold upon which the groups critical for RRE recognition could be displayed, just as the α -helical scaffold of the basic domain of Rev presents the energetically important residues Thr³⁴, Arg³⁵, Arg³⁸, Arg³⁹, Asn⁴⁰, and Arg⁴⁴ to the RNA (Figure 1).^[14] To test this hypothesis, we first assayed a small family of cyclic β -hairpin peptidomimetics (**BIV-1** to **BIV-8**), prepared in earlier work, for their ability to mimic the α -helical Rev protein.^[15]

The RNA-binding sequence of Rev is related to and belongs to the same class of protein (arginine-rich domain) as the Tat proteins of the bovine immunodeficiency virus (BIV) and HIV. Recently, we successfully mimicked BIV Tat using macrocyclic peptidomimetics containing a hairpin-inducing D-Pro-L-Pro template (Figure 2, left).^[15,16] Owing to the high similarity of the arginine-rich regions of Tat and Rev proteins, the same BIV Tat-derived β -hairpin peptidomimetics were tested for in vitro binding to RRE, using polyacrylamide electrophoretic mobility shift assays (EMSA).^[15,16] The Rev-derived peptide (Rev34–50) was used as a positive control in all binding assays.^[4] The dissociation constant (K_d) of this Rev-derived peptide was determined by EMSA to be approximately 100 nM (Figure 3) in the presence of a large excess (280 $\mu\text{g mL}^{-1}$) of *E. coli* tRNA that was used as a control for nonspecific binding. In the absence of excess

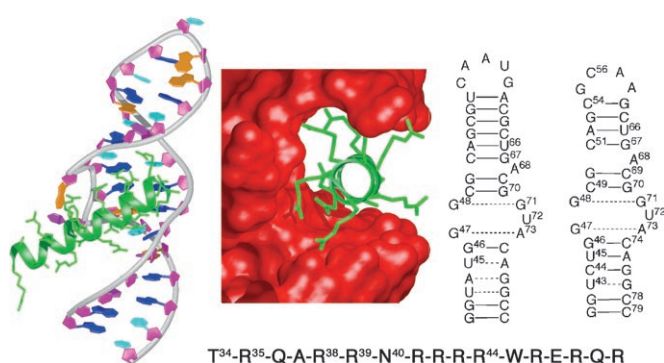


Figure 1. Left: X-ray structure of Rev (green) bound to HIV-1 RRE IIb (prepared from protein data base (PDB) number: 1ETF); middle: key residues in Rev (green) bound in the major groove of the RRE (red surface); right: secondary structure of RRE IIb Rev binding site, used in EMSA studies and the secondary structure of a modified RRE IIb used for NMR spectroscopic studies; bottom: sequence of the arginine-rich RNA binding domain (Rev34–50) of HIV-1 Rev protein.

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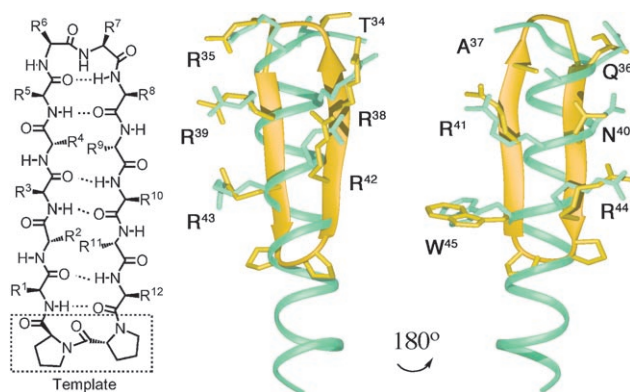


Figure 2. Left: a prototypical 2:2 β -hairpin mimetic (residues 1–12 with a D-Pro-L-Pro template); middle and right: a model β -hairpin (yellow) superimposed (the C(β) atoms of the side chains shown were used as the anchor points for the superimposition) on the Rev helical peptide (cyan). The β -hairpin backbone can be used as a scaffold to pre-organize side chains in a geometry similar to that seen in the helical peptide.

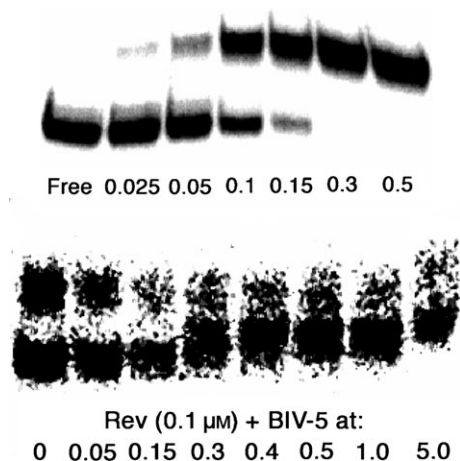


Figure 3. Top: binding assays (EMSA) for the wild-type Rev peptide to RRE (1 nM); bottom: inhibition of Rev peptide (0.1 μ M) from a preformed complex with RRE (1 nM) by peptidomimetic **BIV-5** (all concentrations in μ M). The lower band corresponds to free RNA, the upper to bound RNA.

tRNA, the K_d of Rev–RRE binding was about 20 nM, which is comparable to previous reports.^[4]

Among the peptidomimetics tested in the same way, **BIV-5** exhibited the highest affinity for RRE with a K_d of 300 nM in the presence of tRNA (data not shown), only two-times higher than that measured for the wild-type Rev peptide under the same conditions (Table 1). Two additional molecules, **BIV-2** and **BIV-7**, were also found to bind to the RRE with K_d values in the low micromolar range (data not shown). The other mimetics failed to bind even at high micromolar concentrations. It was shown earlier that **BIV-2** was the tightest-binding ligand for BIV TAR ($K_d = 0.15 \mu$ M), followed by **BIV-5** ($K_d = 1–2 \mu$ M), whereas for **BIV-7** no binding to TAR was detected.^[15] **BIV-5**, was also tested in an inhibition assay to measure its ability to displace Rev peptide in a preformed Rev–RRE complex. **BIV-5** was found to displace

Table 1: Sequences and binding affinities of peptides **BIV-5** and **R-01** to **R-28** to RRE, as determined in EMSA.^[a]

Mimetic	Position												K_d [μ M]
	1	2	3	4	5	6	7	8	9	10	11	12	
BIV-5	R	R	G	T	R	G	K	R	R	I	G	R	0.3
R-01	W	R	R	R	A	T	^D R	Q	R	N	R	R	1
R-02	W	R	R	R	A	P	^D R	Q	R	N	R	R	1
R-03	W	R	R	R	G	P	^D R	Q	R	N	R	R	0.5
R-04	W	R	R	R	V	P	^D R	Q	R	N	R	R	1
R-05	W	R	R	R	A	G	^D R	Q	R	N	R	R	0.4
R-06	W	R	R	R	A	G	K	Q	R	N	R	R	0.3
R-07	W	R	R	R	A	T	^D R	G	R	N	R	R	0.3
R-08	W	R	R	R	A	P	^D R	G	R	N	R	R	0.5
R-09	W	R	R	R	G	P	^D R	G	R	N	R	R	0.5
R-10	W	R	R	R	G	T	^D R	Q	R	N	R	R	0.7
R-11	W	R	R	R	V	G	^D R	Q	R	N	R	R	0.5
R-12	W	R	R	R	A	S	^D R	Q	R	N	R	R	0.4
R-13	W	R	R	R	A	K	G	Q	R	N	R	R	0.2
R-14	W	R	L	R	A	K	G	Q	R	N	R	R	nd
R-15	W	R	Q	R	A	K	G	Q	R	N	R	R	nd
R-16	W	R	R	R	A	K	G	Q	R	N	R	V	0.5
R-17	W	R	L	R	A	^D R	G	Q	R	N	R	R	0.4
R-18	W	R	L	R	A	K	K	Q	R	N	R	R	0.3
R-19	W	R	K	R	A	G	K	Q	R	N	R	R	0.4
R-20	W	R	K	R	A	K	G	Q	R	N	R	R	0.2
R-21	W	R	R	R	A	^D R	G	Q	R	N	R	R	0.5
R-22	W	R	L	R	A	^D R	G	K	R	N	R	R	0.25
R-23	W	R	R	R	A	G	R	Q	R	N	R	R	0.15
R-24	R	R	L	R	A	^D R	G	Q	R	N	R	R	0.1
R-25	R	R	L	R	A	K	G	Q	R	N	R	R	0.1
R-26	R	V	R	R	A	K	G	Q	R	N	R	R	0.3
R-27	R	C	R	R	A	K	G	Q	R	R	C	R	0.1
R-28	K	R	Q	R	T	K	G	R	R	L	O	R	0.1

[a] Positions 1–12 refer to residues 1–12 as mounted on the D-Pro-L-Pro template (Figure 2, left). The dissociation constants (K_d) were determined in the presence of a large excess (280 μ g mL^{−1}) of *E. coli* tRNA that was used as a control for nonspecific binding. Binding reactions were conducted in Tris-HCl buffer (50 mM, pH 8.0), KCl (50 mM), 1,4-dithiothreitol (DTT; 200 mM) and Triton X-100 (0.05 %). nd = no binding detected.

the Rev peptide from the Rev–RRE complex with an IC₅₀ of about 300 nM (Figure 3), which is similar to the estimated K_d for direct binding to the RRE.

To establish that binding of **BIV-5** occurred at the Rev binding site on the RRE RNA, the **BIV-5**–RRE interaction was studied by NMR spectroscopy. Resonances in 2D TOCSY spectra in the free RNA were derived from the known assignment of RRE RNA.^[17] From the TOCSY spectrum (in which pyrimidine H5–H6 resonances could be selectively identified, Figure 4), it was found that mainly resonances at or near the purine-rich internal RNA loop are shifted in the presence of a 1:1 ratio of **BIV-5**. In particular, the U72 resonance was greatly shifted downfield in the presence of **BIV-5**, in agreement with the structural changes observed upon Rev binding when U72 becomes bulged-out. Moreover, the C69 resonance disappeared and the C51, C74, and U66 resonances were shifted, further indicating the specific binding of **BIV-5** to the internal loop region. In contrast, resonances arising from the apical loop and the stem region close to the apical loop and from the 5' and 3' ends either remain unaffected or are shifted slightly (Figure 4).

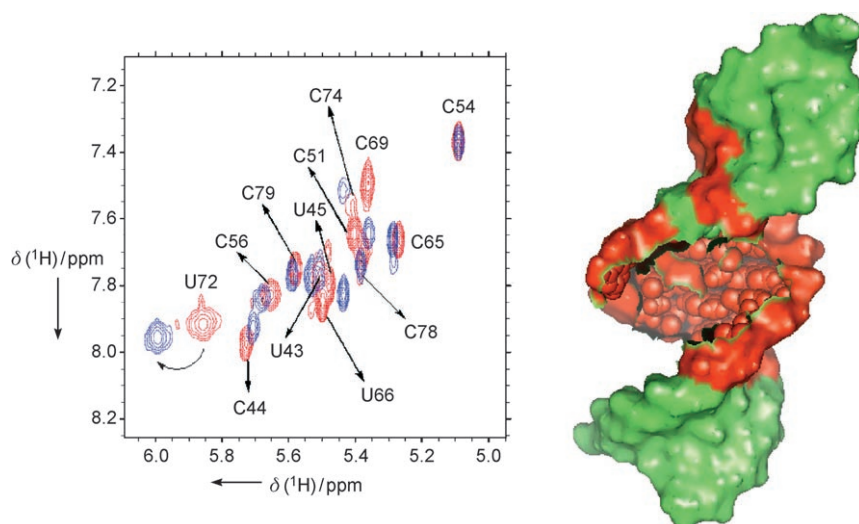


Figure 4. Left: superimposition of TOCSY spectra of free RRE-RNA in red (sequence shown in Figure 1 top, right) and 1:1 **BIV-5**:RRE complex in blue. Resonances at or near the purine-rich internal loop are affected by peptide binding, while the rest of the RNA is not. Homonuclear ^1H TOCSY and NOESY experiments of the free RNA and **BIV-5**:RRE complex were recorded at 750 MHz (Bruker DMX-750) at RNA concentrations of 0.5–1.5 mM in D_2O at pH 6.6 in phosphate buffer; right: schematic representation of the region of RRE affected by peptide binding (red) and the region that remained unaffected (green).

These data are consistent with **BIV-5** binding to an RNA structure which is similar to the RNA structure recognized by Rev.^[6]

These promising results prompted the synthesis of a second library of β -hairpin peptidomimetics (**R-01** to **R-28**, Table 1), which were made to test a new design concept. This concept is based on the premise that, in the Rev-RRE complex, the key RNA-interacting side chains in Rev are displayed around almost the entire circumference of the Rev α -helix (Figure 1, middle). Superimposing a model β -hairpin (yellow, in Figure 2, right) on this Rev helical peptide, revealed that the side chains of residues 1, 12, 3, 10, 5, and 8 on one face of the 12-mer hairpin have approximately the same orientations as the side chains of residues W45, R44, N40, R41, A37, and Q36 in Rev. In a similar way, residues R35, T34, R38, R39, R42, and R43 in the Rev helix might be spatially mimicked by corresponding residues at positions 6, 7, 9, 4, 2, and 11, respectively, on the other face of the hairpin. Moreover, the mimicry appears optimal when Arg7 in the hairpin (which should mimic Arg35 in Rev) has the D-configuration (consistent with this hypothesis, switching from D- to L-Arg reduced activity). Thus, the first mimetic (**R-01**, Table 1) has the sequence $\text{W}^1\text{R}^2\text{R}^3\text{R}^4\text{A}^5\text{T}^6\text{R}^7\text{Q}^8\text{R}^9\text{N}^{10}\text{R}^{11}\text{R}^{12}$ mounted upon the D-Pro-L-Pro template (Figure 2, left). A series of related mimetics (**R-02** to **R-23**) were also studied, which differ in sequence mainly at the tip of the β -hairpin, in an effort to identify a motif that promotes a stable β -turn in this region.

The peptidomimetics **R-01** to **R-23** were synthesized, purified (see Supporting Information) and assayed for binding to RRE by EMSA. Their K_d values measured in the presence of excess tRNA are shown in Table 1. With the exception of peptides **R-14** and **R-15** that did not retain

binding, all the other peptides exhibited a strong affinity to RRE under the conditions tested. NMR spectroscopic studies on several of these mimetics in unbound form in aqueous solution, however, suggested that few regular β -hairpin structures were present, that is, that they are disordered. For example, $^3J(\text{NH}, \text{C}(\alpha)\text{H})$ values were all in the range 6–8 Hz, and few characteristic long-range NOEs could be detected. Although the binding assays show high-affinity binding to the RNA, the lack of stable hairpin structure in these free ligands in solution prompted further studies aimed at stabilizing the hairpin fold.

The final group of mimetics tested (**R-24** to **R-28**) combine features of **BIV-5** and **R-01** to **R-23**, and include a K^6G^7 motif at the hairpin tip which is expected to promote a stable β -turn, as seen in a closely related TAR-binding mimetics.^[18] In addition, they have a disulfide bridge at a non-hydrogen-bonding position in **R-27**, to further stabilize a β -

hairpin structure. Peptides **R-24**, **R-25**, **R-27**, and **R-28** were found to bind to RRE with $K_d \approx 0.1 \mu\text{M}$, similar to the Rev-derived peptide (see Figure 5 for representative data for **R-**

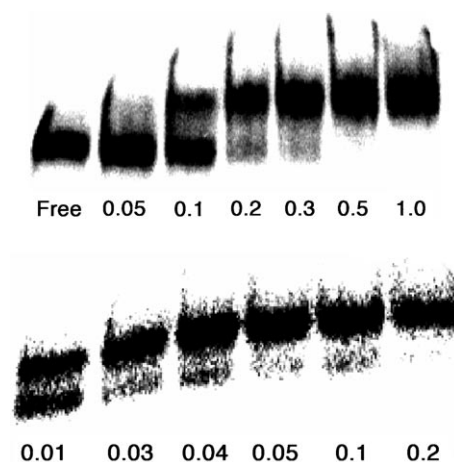


Figure 5. Top: binding assay of **R-24** to RRE (1 nM) in the presence of excess tRNA; bottom: binding assay of **R-24** to RRE (1 nM) in the absence of tRNA (concentrations of **R-24** in μM).

24). When these molecules were assayed in the absence of excess tRNA, a K_d of 10 nM was observed for **R-24** (Figure 5) and **R-25**, whereas **R-27** showed a K_d of 1–2 nM. The affinity of this latter peptide for the RNA is thus significantly stronger than that of the Rev-derived α -helical peptide.

NMR spectroscopic studies on **R-27** free in solution revealed a relatively stable β -hairpin structure for this peptide (see Supporting Information), in which the two strands of antiparallel β -sheet are connected by a type I' turn at K^6G^7 . In

particular, cross-strand NOEs characteristic of a β -hairpin are now observed, and the $^3J(\text{NH}, \text{C}(\alpha)\text{H})$ values are predominantly greater than 8 Hz for residues within the β -strands. Thus the disulfide cross-link is clearly important in stabilizing the regular β -hairpin structure.

A further key issue in RNA ligand design is the specificity of RNA binding, which could clearly be influenced by the flexibility of the ligand as well as by the overall positive charge of this class of peptides. As a very stringent test of binding specificity to RRE, we also evaluated binding of the mimetics to the structurally very similar HIV TAR RNA. As a control, we also tested binding of a peptide containing 7 consecutive Arg residues ($\text{Ac-Arg}_7\text{-NH}_2$), which is expected to interact strongly but nonspecifically with both RNAs, mainly through electrostatic interactions. Consistent with this hypothesis, and demonstrated by EMSA, the Arg_7 peptide binds nonspecifically with a K_d of approximately 2 nM to both HIV TAR and RRE (Table 2). Among the ligands tested **R-**

Table 2: Binding affinities (in μM) of selected peptides to HIV RRE and TAR RNAs.^[a]

Mimetic	K_d (RRE)	K_d (TAR)
R-24	0.010	0.005
R-25	0.010	0.010
R-26	0.005	0.010
R-27	0.002	0.100
R-28	0.002	0.010
Arg_7	0.002	0.002

[a] The affinities were determined by EMSA in the absence of tRNA.

26 and **R-28** discriminate only modestly (less than tenfold) between RRE and TAR. However, **R-27** discriminates by approximately 50-fold between the two closely related RNA structures. Given how closely related these RNA targets are, and the relatively poor specificity typically seen in the interactions of other RNA-binding molecules with more diverse RNA targets, this result represents a notable level of RNA-binding specificity.

The structural and binding data together provide strong evidence supporting our hypothesis that this family of β -hairpin peptidomimetics can mimic the α -helical Rev peptide in its binding to RRE RNA. Although we cannot yet rationalize the quantitative effects of individual side-chain substitutions on the binding energy, we were able to transplant the molecular interactions observed on the α -helical Rev-derived peptide onto a β -hairpin scaffold and very rapidly discover ligands that are more potent than the Rev

protein. Furthermore, the results suggest that not just the number of positively charged groups, but also their relative orientations, as determined by the flexibility of the β -hairpin mimetic, have a decisive influence on the affinity and specificity of RNA binding. These important conclusions merit further investigation, not least since this family of mimetics represents a relatively new class of RNA-binding molecules, with potential for development into novel drugs against HIV.

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- [1] M. Emerman, R. Vazeux, K. Peden, *Cell* **1989**, 57, 1155.
- [2] M. H. Malim, J. Hauber, S. Y. Le, J. V. Maizel, B. R. Cullen, *Nature* **1989**, 338, 254.
- [3] D. A. Mann, I. Mikaelian, R. W. Zimmel, S. M. Green, A. D. Lowe, T. Kimura, M. Singh, G. Butler, M. Gait, J. Karn, *J. Mol. Biol.* **1994**, 241, 193.
- [4] J. Kjems, B. J. Calnan, A. D. Frankel, P. A. Sharp, *EMBO J.* **1992**, 11, 1119.
- [5] J. L. Battiste, *Biochemistry* **1994**, 33, 2741.
- [6] J. L. Battiste, H. Mao, N. S. Rao, R. Tan, D. R. Muhandiram, L. E. Kay, A. D. Frankel, J. R. Williamson, *Science* **1996**, 273, 1547.
- [7] Y. Gosser, T. Hermann, A. Majumdar, W. Hu, R. Frederick, F. Jiang, W. Xu, D. Patel, *Nat. Struct. Biol.* **2001**, 409, 146.
- [8] J. A. Ippolito, T. A. Steitz, *J. Mol. Biol.* **2000**, 295, 711.
- [9] X. Ye, A. Gorin, A. D. Ellington, D. J. Patel, *Nat. Struct. Biol.* **1996**, 3, 1026.
- [10] D. W. Boykin, J. Kumar, J. Spychala, M. Zhou, R. J. Lombardy, J. D. Wilson, C. C. Dykstra, S. K. Jones, J. E. Hall, R. R. Tidwell, C. Laughton, C. M. Nunn, S. Neidle, *J. Med. Chem.* **1995**, 38, 912.
- [11] R. L. Chapman, T. B. Stanley, R. Hazen, E. P. Garvey, *Antiviral Res.* **2002**, 54, 149.
- [12] E. S. DeJong, C.-E. Chang, M. K. Gilson, J. P. Marino, *Biochemistry* **2003**, 42, 8035.
- [13] R. Fasan, R. L. A. Dias, K. Moehle, O. Zerbe, J. W. Vrijbloed, D. Obrecht, J. A. Robinson, *Angew. Chem.* **2004**, 116, 2161; *Angew. Chem. Int. Ed.* **2004**, 43, 2109.
- [14] R. Tan, A. D. Frankel, *Biochemistry* **1994**, 33, 14579.
- [15] Z. Athanassiou, R. L. A. Dias, K. Moehle, N. Dobson, G. Varani, J. A. Robinson, *J. Am. Chem. Soc.* **2004**, 126, 6906.
- [16] T. C. Leeper, Z. Athanassiou, R. L. A. Dias, J. A. Robinson, G. Varani, *Biochemistry* **2005**, 44, 12362.
- [17] J. L. Battiste, R. Tan, A. D. Frankel, J. R. Williamson, *J. Biomol. NMR* **1995**, 6, 375.
- [18] Z. Athanassiou, K. Patora, R. L. A. Dias, K. Moehle, J. A. Robinson, G. Varani, *Biochemistry* **2007**, 46, 741.